

行政院國家科學委員會專題研究計畫 期末報告

細胞核基質蛋白 Matrin 3 (MATR3)在人類 D 型肝炎病毒 RNA 的合成過程中的重要性

計畫類別：個別型
計畫編號：NSC 101-2320-B-040-006-
執行期間：101年08月01日至102年07月31日
執行單位：中山醫學大學醫學研究所

計畫主持人：許國堂

計畫參與人員：博士班研究生-兼任助理人員：丘翎燕

公開資訊：本計畫涉及專利或其他智慧財產權，2年後可公開查詢

中華民國 102年10月17日

中文摘要： 人類的 D 型肝炎病毒(HDV)具有一個環狀單股的核酸(RNA)做為它的基因體。因此，使用其 RNA 為鑄模來做為合成新基因體之用。到目前為止，HDV 的基因體中，只有發現一個基因。這個基因的產物就是 D 型肝炎病毒的抗原(HDAg)。細胞核酸聚合酶的 pol II 可能是主要的核酸聚合酶用來轉錄 HDAg-mRNA，可是 pol II 只能使用 DNA 為鑄模，如何讓它扭轉成以 RNA 為鑄模的專一性轉錄，還未有任何合理的解釋。假如 pol II 可以辨識 HDV 的 RNA 並做為鑄模進行轉錄工作，則我們可以合理推測其中必須要有中間者才可以把 pol II 的工作羣和 HDV 的 RNA 銜接起來。這樣或許是 pol II 可以把 HDV 的 RNA 拿來進行轉錄工作的一種步驟。所以我們在此計畫中推論 MATR3 應該是經過 D 型肝炎病毒的抗原來參與轉錄 HDAg-mRNA 的重要因素。為了找出細胞內可能參與 HDAg 基因轉錄的蛋白質，我們已經建立了會表現 HDAg 的 Huh7 細胞株來研究會被 HDAg 向上調控的蛋白質，並且經過蛋白質體學之二維電泳分析和液相層析串聯質譜的分析之後，找到了在細胞核液中的 Matrin3。

Matrin3 是細胞高度保留的細胞核內基質蛋白，和細胞膜內膜可以形成細胞核內骨架來參與染色質編排、DNA 修復、轉錄和 RNA 修飾。Matrin3 具有核定位訊息序列(NLS)，兩個鋅指 DNA 結合區(Zinc finger)預計會和 DNA 結合。同時它也有兩組 RNA 辨識模體(RRM)預計會和 RNA 結合。因為 Matrin3 是 pol II 的工作羣而且會和細胞核內的修飾過的 RNA 結合，所以在此部份之後續計畫，我們假設 pol II 的工作羣中的 Matrin3 是經由 HDAg 才能用 HDV 的 RNA 拿來進行轉錄工作。我們並且從微矩陣基因晶片技術的分析也看到 MATR3 和 p54nrb/NonO 的表達量還是有增加的。也觀察到在表現 HDAg 的 Huh7 細胞株中 Matrin3 和它的伙伴 p54nrb/NonO 的表達量是有增加的。我們將使用基因高度表達和 RNA 干擾抑制方法對 Matrin3 在複製 HDV 時的角色進行研究。我們已經把 MATR3 和 p54nrb/NonO 的表達載體建立起來。因為我們已經建立了可以完全不用 DNA，可以很接近 HDV 在細胞內複製的狀態的模式進行實驗，所以我們可以專一性地辨識細胞的 HDV 核酸聚合酶(RdRP)的工作羣。是以我們將有機會瞭解到 pol II 工作羣如何和 HDV RNA 進行作用，這對於醫學和生物學上都可以有相當的影響性。

中文關鍵詞： D 型肝炎病毒； D 型肝炎病毒抗原； 蛋白質體學； Matrin3； RNA 為鑄模的核酸聚合酶； 微矩陣基因晶片

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stranded circular RNA genome that replicates by RNA-directed RNA synthesis. The virus encodes only a single protein product, the hepatitis delta antigen (HDAg). It has been proposed that transcription of HDAg-mRNA is mediated by RNA polymerase II, a cellular DNA-directed RNA polymerase. Due to the lack of natural infection for in vivo model and a reliable in vitro transcription assay system, the hunt for the human cellular HDV RNA-directed RNA polymerase (RdRP) is hampered. If genomic of HDV RNA can be recognized by the DNA-dependent pol II, there must be a switch exist that connects the pol II transcriptional machinery and HDV RNA to synthesize mRNA of HDAg. To identify cellular proteins that involved in HDAg transcription, we have analyzed the nuclear lysate of Huh-7 stable clones that expressing HDAg by proteomics on two-dimensional polyacrylamide gel electrophoresis to detect the HDAg-upregulated proteins. From these experiments, Matrin3 was further identified by LC-MS/MS analysis.

Matrin3 is a highly conserved, inner nuclear matrix protein of 125 kDa. Nuclear matrix proteins bound to the inner nuclear membrane form a skeletal nuclear framework with roles in chromatin organization, DNA repair, transcription, and RNA processing. Matrin3 contains a bipartite nuclear localization signal (NLS), two zinc finger domains predicted to bind DNA, and two RNA recognition motifs (RRM). We further postulated that Matrin3 may connect the DNA-dependent pol II machinery via HDAg to synthesize the HDAg-mRNA. We also applied DNA microarray technology to analyze gene expression altered by HDAg and found both Matrin3 and p54nrb/NonO, were upregulated. The expressions of Matrin3 and its partner, p54nrb/NonO, were upregulated in small-HDAg expressing Huh7 cells that examined by immunoblotting.

To characterize the role of Matrin3 plays in HDV RNA synthesis, we went on to study the essential role of Matrin3 in HDV replication by gene overexpression and RNA interference. We have cloned and constructed the expression vectors of Matrin3 and p54nrb/NonO. Since we already established a cDNA-free HDV replication

system in cellular model, we are able to specifically analyze the cellular RdRP machinery from human cell that responsible for HDV RNA synthesis. How to switch the pol II machinery from DNA-dependence to recognize HDV RNA as the template for RNA synthesis is an important issue in the fields of biology and medicine.

英文關鍵詞： Hepatitis delta virus； Hepatitis delta antigen；
Proteomics； Matrin 3； RNA-directed RNA polymerase；
DNA microarray

行政院國家科學委員會補助專題研究計畫
畫

期中進度報告
期末報告

(計畫名稱)

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The role of an inner nuclear matrix protein Matrin 3 (MATR3) in human hepatitis delta virus RNA synthesis

計畫類別：個別型計畫 整合型計畫

計畫編號：NSC 101-2320-B-040-006-

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執行機構及系所：中山醫學大學醫學研究所

計畫主持人： 許國堂

共同主持人：

計畫參與人員：兼任助理人員：丘翎燕

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涉及專利或其他智慧財產權，一年 二年後可公開查詢

中 華 民 國 102 年 10 月 30 日

中文摘要及關鍵詞 (keywords):

D 型肝炎病毒; D 型肝炎病毒抗原; 蛋白質體學; Matrin 3; RNA 為鑄模的核酸聚合酶; 微矩陣基因晶片

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前言：

Background of Hepatitis delta virus (HDV)

HDV life

HDV has a circular, single-stranded RNA genome of 1,700 nucleotides, with about 70% self-complementarily (Kos et al., 1986; Kuo et al., 1988a; Makino et al., 1987; Wang et al., 1986). This genome is thought to be replicated by the host RNA polymerase II through a double-rolling-circle mechanism (Chen *et al.*, 1986; Fu & Taylor, 1993; Lee *et al.*, 1993). Replication of HDV genome is independent to HBV infection (Chen et al., 1986). Once enter the cell, HDV can produce linear multimers of both genomic and antigenomic RNAs, both of which possess a self-cleaving activity that processes the RNAs to unit length (Kuo et al., 1988b; Sharmeen et al., 1988; Wu et al., 1989). There are two detectable HDV antigens (HDAg) encoded by a single open reading frame on the antigenomic strand (Bergmann & Gerin, 1986; Bonino *et al.*, 1986; Pohl *et al.*, 1987; Weiner *et al.*, 1988). The large (L-HDAg) is a result of RNA editing which extends the protein coding sequence of the small antigen (S-HDAg) by 19 residues (Luo et al., 1990).

HDV antigens

The S-HDAg is essential for viral replication and is proposed to facilitate the interactions between the cellular transcription machinery and the viral template (Kuo et al., 1989; Lin et al., 1990). HDAg inhibits cellular RNA polymerase II but not the pol I and pol III *in vivo* with a CAT reporter assay and *in vitro* with the purified recombinant HDAg have been shown (Lo et al., 1998), suggesting that mass amount of HDAg capable interact with pol II specific transcriptional component. The central domain of S-HDAg, which is involved in binding to the viral genome, contains two arginine-rich motifs bracketing a leucine zipper (Lee et al., 1993). The nuclear import of HDV capsid protein may be the first biological function of S-HDAg that leads to the replication of HDV RNA in the nuclei of infected cells (Chou et al., 1998). L-HDAg appears later in infection, suppresses replication and initiates viral packaging (Chang et al., 1991; Chao et al., 1990). We have reported that HDV replication was associated with epigenetic regulation resulted in histone hyperacetylation and clusterin induction. Thus, both isoforms of the HDAg may act as co-activators of cellular gene expression (Liao *et al.*, 2009).

HDV-RNAs

Three major stable RNA species are thought to be involved in the replication of the HDV RNA genome. (a) The genomic 1,679-nucleotide circular single-stranded RNA. (b) Its exact complement, the antigenome, and (c) an 800-nucleotide polyadenylated RNA that is least abundant for the mRNA of HDAg (Gudima *et al.*, 1999; Perrotta & Been, 1990; Taylor, 1990). It has been proposed that the 800-nt mRNA is continuing synthesized throughout HDV replication that is not suppressed by the translated HDAg as previously reported (Modahl & Lai, 1998).

HDV RNA editing

During HDV genome replication in the nucleus, some of the new antigenomic RNAs get edited by an enzyme of the class known as adenosine deaminase acting on RNA (ADAR). The small form of ADAR1, is considered to be the specific enzyme that acts on HDV RNA (Wong & Lazinski, 2002). It changes adenosine to inosine. Editing occurs specifically on adenosine 1012 (the amber/W site) of the

antigenome (Casey & Gerin, 1995) and changes the UAG (amber) stop codon of S-HDAg to a UGG (Trp) codon, which allows for the C-terminal addition of the 19 to 20 amino acids unique to L-HDAg. This edited change is processed by subsequent rounds of RNA-directed RNA synthesis, as adenosine to guanosine. Typically, 20-50% of antigenomic RNA was edited *in vitro* and *in vivo* (Polson et al., 1996) and is suppressed by S-HDAg (Polson et al., 1998).

Systems have been used to replicate HDV in cell lines

HDV replication products have been obtained by several transfection approaches that bypass the processes of virus attachment and penetration: (A) Transfection with a cDNA plasmid containing three tandem head-to-tail inserts of the HDV genome under the control of SV-40 promoter (Kuo et al., 1989) (B) Transfection of *in vitro* synthesized genomic RNA into S-HDAg stable expressed fibroblasts (Glenn et al., 1990). (C) Co-transfection of *in vitro* synthesized HDV RNA and mRNA of S-HDAg into Huh-7 cells (Modahl & Lai, 1998). (D) Transfection of HDV RNA and recombinant S-HDAg ribonucleoprotein complex (RNP) into Huh-7 cells (Dingle *et al.*, 1998; Sheu & Lai, 2000). All of the four approaches require the presence of the S-HDAg for HDV replication. We have established the system of (B) that replicates *in vitro* synthesized genomic RNA in S-HDAg stable expressing Huh-7 cells (Huh-sm). This RNA transfection system is considered as the most reliable system to study the HDV replication (Liao *et al.*, 2012).

Enzymology of RNA-directed RNA replication of HDV

Replication of HDV genomic, antigenomic and message RNAs apparently are mediated by distinct cellular RNA polymerases. The mRNA transcription is inhibited by a low concentration of α -amanitin and could be rescued partially by an α -amanitin-resistant mutant pol II. Therefore, pol II is supposed to synthesize mRNA of HDAg from the HDV RNA genomic template (Modahl et al., 2000). How pol II transcription machineries are converted from DNA-directed synthesis to RNA-directed synthesis is not clear. Our proposal in here is focus at this issue. The synthesis of the anti-genomic HDV RNA was not affected by α -amanitin, therefore, pol II may not responsible for the antigenomic RNA synthesis (Modahl et al., 2000). Moreover, the synthesis of genomic RNA was inhibited by L-HDAg, whereas, anti-genomic RNA synthesis was not affected (Modahl & Lai, 2000). We have shown that only genomic RNA associated with S-HDAg could be used as template for replication, anti-genomic RNA and S-HDAg RNP could not initiate replication (Sheu & Lai, 2000). All these evidences point to that replication and transcription of HDV are mediated by distinct cellular polymerases. Recently, it has been reported that human RNA polymerases II (Greco-Stewart et al., 2007), I and III (Greco-Stewart et al., 2009) interact with the terminal stem-loop region. The RNA pol II forms a preinitiation complex on this region (Abraham & Pelchat, 2008). However, whether the initiation site of 1.7-kb antigenomic or genomic HDV RNA synthesis is identical to that of 0.8-kb mRNA transcription is not known. We demonstrated the nucleotide positions 1582 to 1683 (TR-P1) that located at the terminal stem-loop region are substantial for the maximal promoter activity. Mutations of TR-P1 to P1-m5 completely block the HDV RNA synthesis in Huh-sm cells (Liao *et al.*, 2012). TR-P1 region most likely is the authentic HDV RNA promoter for mRNA transcription.

Study the HDAg regulated proteins by proteomic and immunoprecipitation methods

To identify the changes in the expression profiles of host proteins caused by the presence of the different virus components, Huh7 cells have been transiently transfected with plasmids coding

exclusively for the S-HDAg, L-HDAg, genomic RNA, and antigenomic gRNA, respectively. Total protein extracts were separated by 2-DE and differentially expressed spots were identified by MALDI-TOF followed by database searching (Mota et al., 2008). Although a total of 32 proteins were identified using this approach, whether or not they interact with HDAg and/or affect HDV metabolism was not addressed. Similar approach was applied to Huh7-D12 cells that stably expressing HDV RNPs (Mota et al., 2009) and 23 differentially expressed proteins were identified, of which 15 were down regulated and 8 up regulated in Huh7-D12 cells. A combined proteomic-RNAi screen for host proteins interacting with S-HDAg also has been reported on HEK293 cells that stably express a functional Flag-S-HDAg (Cao et al., 2009). By anti-Flag immunopurification and mass spectrometry analysis, many proteins were identified. Total of 65 proteins were selected to look for whether they would also affect the accumulation of HDV RNA following siRNA knockdown. The results showed most of the proteins tested could affect RNA accumulation.

An alternative approach to search for HDV host factors employed mass spectrometry analysis following UV crosslinking of HeLa nuclear proteins with an RNA corresponding to the right terminal stem-loop domain of HDV genomic RNA (Greco-Stewart et al., 2006). This identified the polypyrimidine tract-binding protein-associated splicing factor (PSF) as a host factor bound to HDV RNA, although PSF functional involvement during physiological HDV replication remains unclear. Additional protein interactions have also been reported (Sikora et al., 2009). The proteins of eEF1A1, p54^{nrb}/NonO, HnRNP-L, GAPDH and ASF/SF2 were co-immunoprecipitated with the right terminal stem-loop domain of HDV genomic RNA in vitro. Those data indicate that HDV RNA associates with RNA-processing pathways and translation machinery during its replication provides new insights into HDV biology. There have been additional reports of host proteins that bind to either HDAg or HDV RNA. PML (Li et al., 2006), nucleolin (Lee et al., 1998), B23 (Huang et al., 2001), and transcription factor YY1 (Huang et al., 2008) have been shown to interact with HDAg and were suggested to affect HDV replication. The Pol I specific factor SL1 (Li et al., 2006) and the negative transcription elongation factor DSIF (Yamaguchi et al., 2001) have also been demonstrated to interact with HDAg, but their effect on HDV replication in vivo is not clear. A double-stranded RNA-activated kinase, PKR, has been shown to bind to S-HDAg (Chen et al., 2002) and HDV RNA (Robertson et al., 1996) (Circle et al., 1997), and affect HDV replication (Chen et al., 2002). Finally, GAPDH was reported to bind to HDV RNA (Lin et al., 2000) and ADAR1 was found to edit HDV antigenomic RNA (Wong & Lazinski, 2002).

Background of MATR3, PSF and p54^{nrb}/NonO

Matrin3

Nuclear matrix is a major component of the cell nucleus (Berezney et al., 1995) and is composed of over 200 proteins (Fey & Penman, 1988). The major proteins of the nuclear matrix include lamins A, B and C, which can form a meshwork within the nucleoplasm (Inagaki et al., 1996): lamin B1, which associates with chromatin at the matrix attachment regions (Inagaki et al., 1996); the nucleolar protein B-23; core heterogeneous nuclear RNPs, and the nuclear matrins (Berezney et al., 1995). The nuclear matrins are components of the internal nuclear matrix and include matrins 1, 3, 4, D, E, F, G, 12 and 13 (Berezney et al., 1995). Of these, matrins D, E, F, G, and 4 have been identified as DNA-binding proteins (Hakes & Berezney, 1991). Matrin3 is a highly conserved, inner nuclear matrix protein of 125

kDa. The primary structure deduced for Matrin3 predicts a bipartite nuclear localization signal (NLS), two DNA binding domains with C₂H₂-type zinc finger domain and two tandem RNA recognition motifs. It has been implicated in the processing of RNAs and retention of hyper-edited RNA within the nucleus (Zhang & Carmichael, 2001). Moreover, Matrin3 has nuclear localization signals and several phosphorylation sites for tyrosine or serine/threonine kinases (Beausoleil et al., 2004). Though its activity and mode of action are still unclear, it appears that Matrin3 serves as a scaffolding molecule that interacts with a number of unique nuclear localized proteins (Malyavantham et al., 2008; Zeitz et al., 2009). A majority of these proteins are involved in RNA metabolism and chromatin remodeling while others function in protein translation, DNA replication/repair and apoptosis.

Polypyrimidine tract-binding protein (PTB)-associated splicing factor (PSF) and p54^{nrb}/NonO

PSF, a 100 kDa polypeptide, was identified and characterized in a complex with PTB (Patton et al., 1993). p54^{nrb} (human) and NonO (mouse) are highly homologous to the C-terminus of PSF (Dong et al., 1993; Yang et al., 1993). Proteomics have identified PSF and p54^{nrb}/NonO in the nucleolus (Andersen et al., 2002) and in association with the nuclear membrane (Dreger et al., 2001). Matrin3 together with the proteins PSF and p54^{nrb}/NonO were implicated in nuclear retention of hyper-edited RNA, which prevents the translation of such RNA (Zhang & Carmichael, 2001). This process occurs in nuclear bodies called paraspeckles that contain PSF and p54^{nrb}/NonO proteins (Bond & Fox, 2009). PSF and p54^{nrb}/NonO form a heterodimer and are involved in various aspects of RNA and DNA metabolism (Shav-Tal & Zipori, 2002) such as transcription (Buxade et al., 2008; Dong et al., 2007; Hata et al., 2008), pre-mRNA 3' processing (Rosonina et al., 2005), transcription termination (Kaneko et al., 2007) and mRNA splicing (Ito et al., 2008; Kameoka et al., 2004). Both PSF and p54^{nrb}/NonO were identified in a biochemical screen for DNA end-rejoining proteins (Bladen et al., 2005). Furthermore, they were shown to facilitate in vitro the DNA binding capability of the KU70/KU80 heterodimer, a component of DNA-PK holoenzyme; and PSF was shown to bind RAD51 and to modulate RAD51-mediated homologous pairing and strand exchange.

方法、結果與討論 (含結論與建議)

This research plan: Investigation of the essential role of Matrin3 in HDV replication by gene overexpression and RNA interference.

Methods:

(1) Proteomics analysis of HDAGs upregulated proteins in Huh7 cells stably expressing S-HDAG or L-HDAG

Results: Matrin3 was further identified by LC-MS/MS analysis.

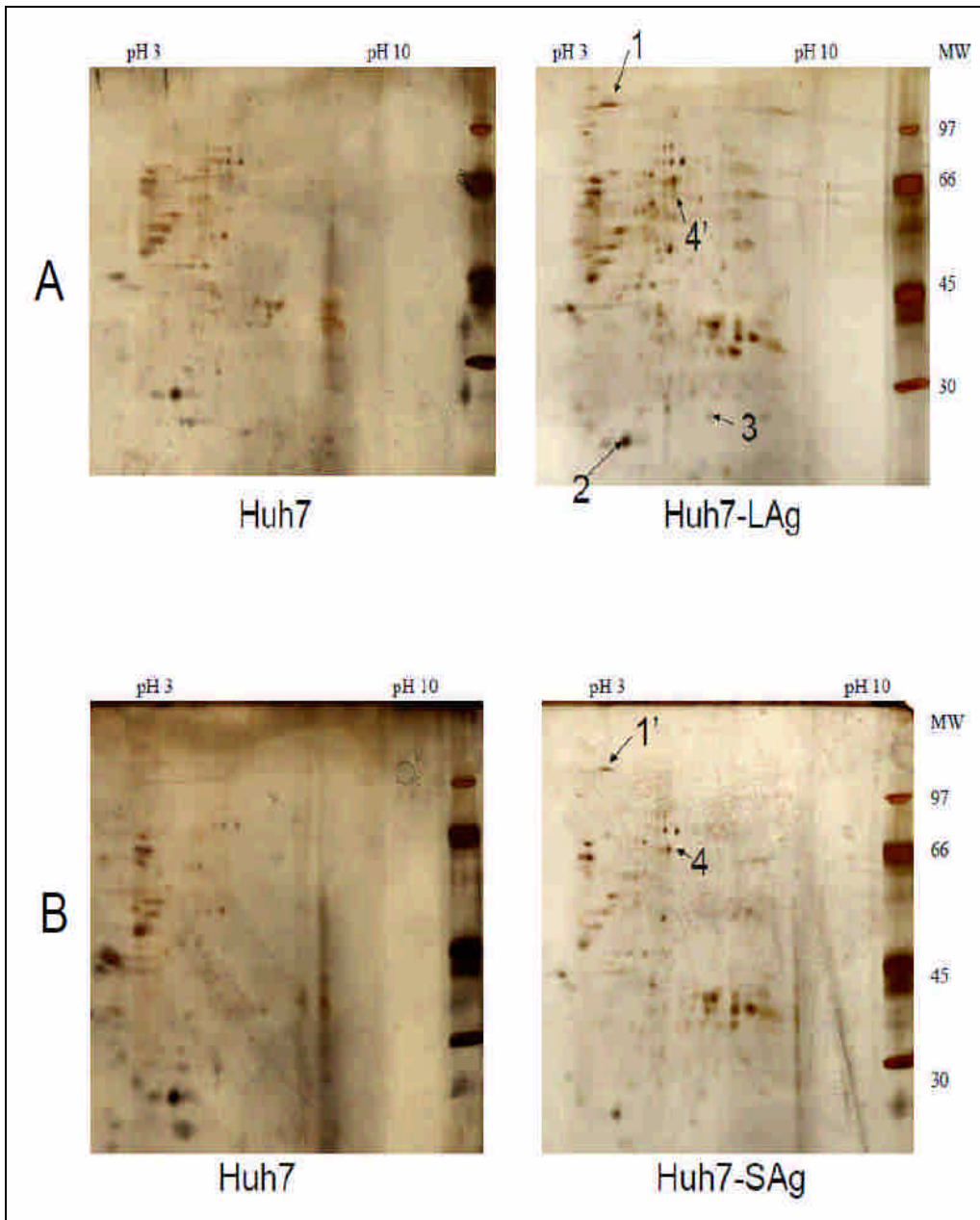


Fig.1 Proteomic analysis of HDAG-regulated cellular proteins by 2D-PAGE and LC-MS/MS. The Huh7 cells that (A) stably expressing L-HDAG (Huh-LAg) and (B) stably expressing S-HDAG (Huh-SAg) were harvested and the nuclear lysates (2 mg) were obtained for proteomic analysis. The upregulated proteins that observed in gels were sliced for LC-MS/MS. Numbers of proteins that were identified are 1: Matrin3; 2: Heat shock protein 27; 3: Glyceraldehydes-3-phosphate dehydrogenase (GAPDH); 4: HnRNP-L. The 1' and 4' indicate similar spots that were not used for analysis and presumed to be the identified one.

Methods:

(2) Analysis of gene expression altered by L-HDAG or S-HDAG with DNA microarray technology.

Results: We found transcription of Matrin3 and p54^{nrb}/NonO was upregulated.

Table 1: The comparison of MATR3, p54^{nrb}/NonO and SFPQ transcription levels in Huh7 cells expressing S-HDAg or L-HDAg.

Gene	Accession No.		Normalized	Normalized
			Huh/SHDAg	Huh/HDAg
MATR3	NM_199189	Matrin3 (MATR3), transcript variant 1, mRNA	1.20x	1.26x
NONO	NM_007363	Homo sapiens non-POU domain containing, octamer-binding (NONO), mRNA	1.37x	1.12x
SFPQ	NM_005066	Splicing factor proline/glutamine-rich (polypyrimidine tract binding protein associated) (SFPQ), mRNA	0.90x	0.99x

Methods:

(3) Examination of the Matrin3 and p54^{nrb}/NonO expressions in Huh7 cells expressing S-HDAg.

Results: The expressions of Matrin3 and p54^{nrb}/NonO, were upregulated in S-HDAg expressing Huh7 cells

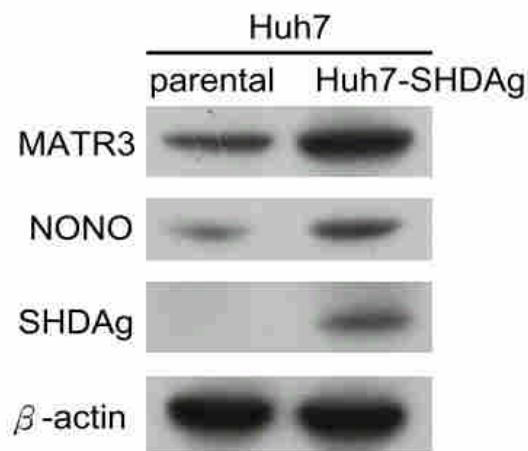


Figure 2. The expression of MATrin3 and p54^{nrb}/NonO were upregulated in Huh7 cell expressing S-HDAg (Huh7-SHDAg) cells that examined by immunoblotting.

Methods:

(4) Construction and transfection of Matrin 3 and p54^{nrb}/NonO expression plasmids into Huh7 cells followed by Western blots analysis

MATR3-F BamHI: 5'-CTCGGATCC ATGTCCAAGTCATTCC
 MATR3-R XhoI: 5'-AGACTCGAG TTAAGTTTCCTTCTTCTGTC

Nono-BamHI-F 5'-CTCCGGATCCATGCAGAGTAATAAAACTT
 Nono-EcoR-R 5'-TGCAGAATTCTTATTATCGGCGACGTTTGTGG

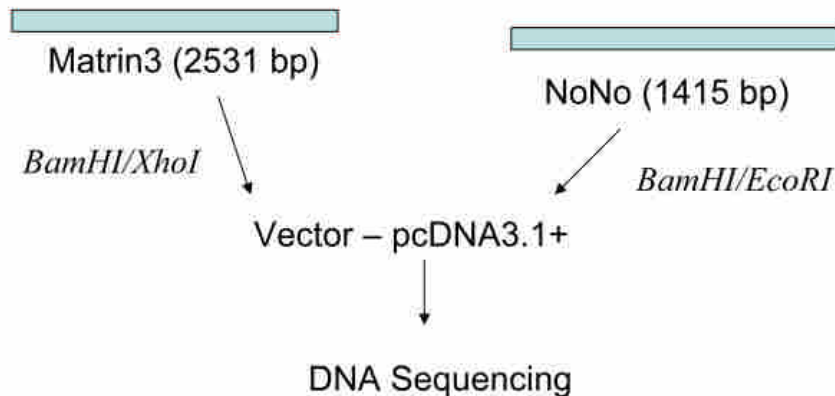


Figure 3. To study the function of Matr3 and p54^{nrb}/NonO, we first construct the expression clone of Matr3 and p54^{nrb}/NonO into expression vector (pcDNA3.1+).

Results: DNA coding Matr3 was obtained by reverse transcription of total RNA isolated from Huh7 cells, by the use of SuperScript III Reverse Transcriptase with primers: MATR3-F BamHI and MATR3-R XhoI. PCR product is cloned into pcDNA3.1+ using *XhoI* and *BamHI* restriction sites. For cloning p54^{nrb}/NonO gene, primers: Nono-BamHI-F and Nono-EcoRI-R were used. The obtained pcDNA-Matr3 and NoNo is propagated in Escherichia coli XL-10, purified with Plasmid Miniprep Kit, and sequenced to confirm the open reading frame. We have successfully obtained Matr3 and p54^{nrb}/NonO in expression vector.

Further experiments to complete the investigation 尚未完成的部份:

(1) The effect of Matr3 and p54^{nrb}/NonO overexpression in HDV RNA replication.

To analyze the effect of Matr3 overexpression in HDV replication, Plasmids of pcDNA-Matr3 and HDV 1.9-kb/pKS will be cotransfected into Huh7 cells with Lipofectamine 2000. At the indicated time points (1, 2, 3, 4 days of posttransfection), the protein lysates and RNAs will be prepared for Western and Northern blots analysis. The data will be compared with the samples that cotransfected with pcDNA empty vector and HDV 1.9-kb/pKS to reveal the effect of Matr3 overexpression by the levels of detected L- and S-HDAg in Western blot. Meanwhile, the levels of monomeric and dimeric HDV RNAs will be compared with the control blots also.

(2) Application of RNA interference of Matr3 followed by Western and Northern blots analysis of HDAGs and HDV RNA.

For silencing the Matr3 gene expression a set of 3 target-specific 20-25 bp siRNAMtr3 (Santa Cruz Biotech., CA) was used: siRNAaMtr3 (411) 5'-GCUACCCAGUCUUUAAGUAtt-3', siRNAbMtr3 (926)5'-CCAUGGAAGUCGUUCUCAAtt-3', siRNAcMtr3 (1504) 5'-CUAGCAGAGUUGUUCACAUtt-3'. They hit all two isoforms of matr3. As a negative control the functional non-targeting siRNAC is used, containing four mismatches for any human, mouse, and

rat gene. The sequence of its sense strand was 5'-UAGCGACUAAACA CAUCAAUU-3'. Synthetic siRNAs (40 nM) and HDV 1.9-kb/pKS plasmids are cotransfected to 50-60% confluent Huh7 cells with Lipofectamine 2000. MATR3 and HDAGs levels in whole cell lysates, and extracted RNA that collected at 1, 2, 3, and 4 days after transfection, will be estimated by Western and Northern blotting. According to our postulation, repression of MATR3 would result in reducing levels of HDAGs and HDV RNAs.

Expected results:

Matrin3 enhances HDV RNA replication if HDAGs and RNA levels are increased in Matrin3 cotransfection sets of samples. Otherwise, the results will indicate the inhibition of HDV replication by Matrin3 overexpression. In contrast, repression of Matrin3 by RNA interference would reduce HDV replication. Matrin3 will be detected by monoclonal antibodies (1:1000) to Matrin3. From the early report, Matrin3 is shown to bind Rev/RRE-containing viral RNA. This binding interaction stabilizes unspliced and partially spliced HIV-1 transcripts leading to increased cytoplasmic expression of these viral RNAs (Yedavalli & Jeang, 2011). Therefore, we postulate that high level of Matrin3 expression would enhance the HDV replication.

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已發表論文：

Hsiao YP, Yang JH, Wu WJ, Lin MH, Sheu* GT. (2013 Oct) E6 and E7 of human papillomavirus type 18 and UVB irradiation corporately regulate interleukin-6 and interleukin-8 expressions in basal cell carcinoma. **Exp Dermatol.** 22(10):672-674. doi: 10.1111/exd.12223, PMID: 24079741 (SCI) (IF=3.578 R/C=6/58=10.3%, **DERMATOLOGY**) (NSC-101-2320-B-040-006)

E6 and E7 of human papillomavirus type 18 and UVB irradiation corporately regulate interleukin-6 and interleukin-8 expressions in basal cell carcinoma

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Abstract: The lack of a human papillomavirus (HPV)-infected skin cancer cell line has hampered the investigation of the interaction of UV and HPV in skin carcinogenesis. We identified a human basal cell carcinoma (BCC-1/KMC) cell line integrated with E6 and E7 genes of high-risk HPV type 18 and demonstrated that repression of E6 and E7 results in proliferation inhibition. Sublethal ultraviolet-B (UVB) irradiation induced the expressions of interleukin-6 (IL-6) and interleukin-8 (IL-8), as well as viral E6 and E7 genes, in BCC-1/KMC cells. When E6 and E7 expressions were inhibited, IL-6/IL-8 expressions were

repressed. Furthermore, IL-6/IL-8 remained inducible by UVB irradiation when E6 and E7 were inhibited. These results indicated that IL-6 and IL-8 can be upregulated by viral E6 and E7 proteins without UVB irradiation. Moreover, chronic exposure to UVB upregulates IL-6 and IL-8 when E6/E7 is induced by UVB.

Key words: BCC – HPV – IL-6 – IL-8 – VUB

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Background

The most commonly diagnosed cancer in the Caucasian population is basal cell carcinoma (BCC), a non-melanoma skin cancer (NMSC) (1,2). UV radiation is considered the most critical risk factor for inducing BCC carcinogenesis (2,3). Therefore, in most cases, BCC develops on chronically sun-exposed skin not only in elderly people (4,5) but also in young- and middle-age individuals (6), while in young adults, it is linked to artificial tanning (7). Fair skin colour is also significantly associated with BCC risk (8). Human papillomaviruses (HPVs) have been found to be associated with NMSCs (9–13). In a previous report, HPV DNA was detected in 27.9% of BCC cases (14). HPV-8 and HPV-18 are the most frequent types of HPVs in immunocompetent patients (15). The association between cutaneous HPV infection and BCC has recently been reported and suggests that β -HPV types play a role in BCC (16). Interestingly, certain types of HPVs, in conjunction with UVB, may be involved in the pathogenesis of NMSCs. The HPV-77 promoter can be stimulated by UV radiation (17). In particular, UVB upregulates interleukin-6 (IL-6) and interleukin-8 (IL-8) to a higher extent in HPV-immortalized cells than in control keratinocytes (18). UVB stimulates keratinocyte cultures to secrete proinflammatory cytokines such as tumor necrosis factor- α (TNF- α), IL-1 β , IL-6, IL-10 and IL-8. The release of these cytokines can trigger a cutaneous inflammatory response to sunburn (19–21). Here, we report a BCC cell line (22) integrated with HPV-18 and further characterize the role of HPV-18 in the regulation of BCC.

Questions addressed

Characterization of the role of E6 and E7 in the interaction with UVB in HPV-18-infected BCC cell line.

Experimental design

Doxycycline-induced shRNA inhibition of E6 and E7 was established in BCC-1/KMC cells. For a detailed description of our methods, please see Data S1.

Results

We first investigated whether the expressions of IL-6, IL-8, E6 and E7 genes are modulated by UVB irradiation in BCC-1/KMC cells. The mRNA levels of IL-6, IL-8, and E6 increased significantly 24 h after UVB (100 J/m²) irradiation, but E6 expressions were dramatically reduced following UVB (200 J/m²) irradiation (Fig. 1a) with significant cell death. These results suggested that UVB irradiation causes IL-6/8 production and E6 expression. Sublethal dose (50 J/m²) of UVB-induced expressions of E6, IL-6 and IL-8, which were further characterized over time (Fig. 1b). Within hours of low-dose UVB irradiation, E6, IL-6 and IL-8 gene expressions gradually increased. The highest levels were recorded 24 h after irradiation.

Next, we applied inducible RNA interference technology to separately target the E6 and E7 genes. After two days of induction by doxycycline (Dox), the E6 mRNAs Sh-E6-1 and Sh-E6-2 were reduced to 30% and 40%, respectively. Similar results were observed in Sh-E7-1 and Sh-E7-2 clones (Figure S1 a, b). When E6 protein level was reduced, p53 protein level increased while E7 protein level decreased (Figure S1 c). Furthermore, when E7 protein level was reduced, E6 protein level decreased, while pRB level increased (Figure S1 d). To further investigate the influence of HPV 18-E6/E7 inhibition, Sh-E6-1 and Sh-E7-1 cells were tested for their ability to proliferate on colony formation assay (Figure S2). The results demonstrated that inhibition of E6 and E7 reduces BCC-1/KMC cell proliferative abilities.

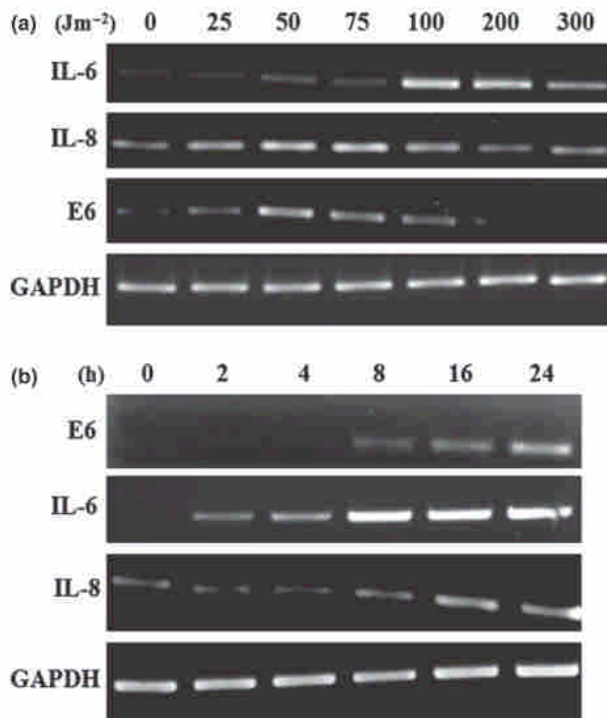


Figure 1. Analysis of E6, E7, IL-6 and IL-8 gene expressions in UVB-radiated BCC-1/KMC cells. (a) The mRNA levels of E6, E7, IL-6 and IL-8 genes were analysed by semiquantitative RT-PCR in BCC-1/KMC cells with low-to-high doses of UVB. IL-6, IL-8 and E6 mRNA expressions were also analysed. (b) The inductions of E6, IL-6 and IL-8 expressions by sublethal dose (50 J/m^2) of UVB were analysed for up to 24 h. GAPDH served as the loading control.

The upregulation of IL-6 expression has been reported in association with the expression of E6 from prototypic skin cancer-associated HPV type 5 (23). Therefore, we further investigated the association between E6/E7 of HPV-18 and IL-6/8 expression. The mRNA levels of IL-6 and IL-8 were reduced after Dox induction of the Sh-E6-1 cells (Fig. 2a). When E7 expression was inhibited by shRNA interference, the mRNA levels of IL-6 and IL-8 were reduced over time following Dox induction of Sh-E7-1 cells (Fig. 2b). Hence, inhibition of E6 and E7 expressions was associated with IL-6 and IL-8 gene repression. To examine the pathway that may be involved in the regulation of IL-6/8 by E6/E7 and UVB, we characterized the expression of IL-6/8 with E6 gene inhibition and UVB irradiation. When compared with the parental BCC-1/KMC cells treated with Dox, under the conditions of Dox-induced E6 inhibition and sublethal dose (50 J/m^2) of UVB irradiation, the ability to upregulate the expression of IL-6/8 was maintained in Sh-E6-1 cells (Fig. 2c).

Conclusions

The human BCC cell line (BCC-1/KMC) was derived from undifferentiated type of facial BCC tumor on thermal traumatic scar. This cell line spontaneously secretes more IL-6 and IL-8 but less

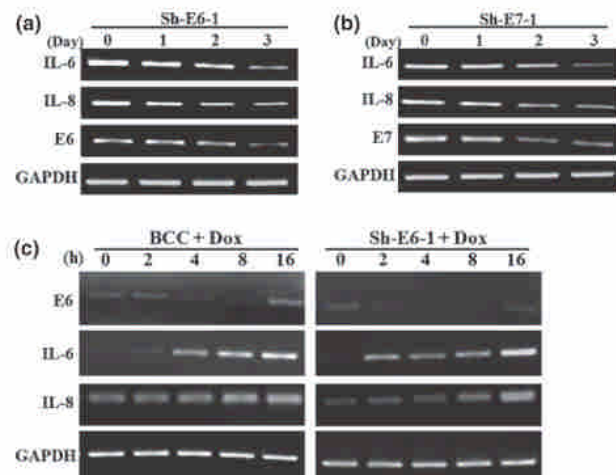


Figure 2. Analysis of IL-6 and IL-8 gene expressions in E6/E7-inhibited cells. (a) The mRNA levels of E6, IL-6 and IL-8 genes were analysed by semiquantitative RT-PCR in Sh-E6-1 cells with Dox induction from 1 to 3 days. (b) Similar conditions were applied to analyse E7, IL-6 and IL-8 in Sh-E7-1 cells. (c) After 3 days of Dox induction, the effects of UVB irradiation (50 J/m^2) on IL-6 and IL-8 in Sh-E6-1 cells were determined. The mRNA levels of E6, IL-6 and IL-8 genes were analysed at the indicated times by semiquantitative RT-PCR after irradiation.

IL-1 than keratinocytes (22), regardless of HPV-18 infection status. The E6 and E7 oncoproteins may contribute to tumor initiation and play important roles in malignant progression through the induction of genomic instability and other mechanisms (24). The complete loss of p53 was shown to result in upregulated expression of the hedgehog pathway, with activated pathway being a prerequisite for the development of BCC (25). Our data may indicate that loss of p53 by E6 of HPV-18 promotes the development of advanced-stage BCC even without UVB exposure.

HaCat keratinocytes transduced with both E6 and E7 genes of beta-HPV types do not demonstrate any inhibition of UVB-induced apoptosis (26). In our study, repression of E6/E7 expression concomitantly reduced the IL-6 and IL-8 expressions and cell proliferation. Sublethal doses of UVB not only upregulated proinflammatory cytokines, but also stimulated the expression of E6/E7. Even without UVB exposure, HPV E6/E7 is still capable of stimulating proinflammatory cytokines. This is the first report to demonstrate that E6/E7 of HPV-18 is essential for BCC-1/KMC cell line survival and cooperates with UVB to upregulate proinflammatory cytokines. Therefore, these results further support the aetiological role of HPV infection in skin cancer.

Acknowledgements

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Conflict of interests

The authors have declared no conflicting interests.

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Supporting Information

Additional Supporting Information may be found in the online version of this article:

Figure S1. Inhibition of E6 and E7 expressions by inducible shRNA technique alters p53 and pRb protein levels.

Figure S2. Characterization of the effects of viral E6 and E7 gene inhibition on cell proliferation.

Data S1. Materials and methods.

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Letter to the Editor

Effects of a topical aqueous oxygen emulsion on collagen deposition and angiogenesis in a porcine deep partial-thickness wound model

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Abstract: A porcine deep partial-thickness wound model was used to evaluate the effects of a newly developed topical aqueous oxygen emulsion (TOE) on wound repair. The wounds were treated with TOE, which contains super-saturated oxygen or vehicle control. Semiquantitative immunofluorescent staining was performed to examine protein production for type I and type III collagen and vascular endothelial growth factor (VEGF). Immunofluorescent staining revealed higher protein levels of type I and type III collagen and VEGF in the TOE treatment group. Histological analysis also revealed improved angiogenesis and granulation tissue formation with topical TOE treatment and was

consistent with the protein expression. In addition, the histology examination demonstrated faster epithelialization in wounds treated with TOE. The study suggests that sustained high levels of oxygen released by TOE may promote the process of wound repair through increasing collagen deposition and angiogenesis as well as stimulating epithelialization.

Key words: collagen – granulation tissue formation – partial-thickness wound – topic aqueous oxygen emulsion – VEGF

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Background

Oxygen plays a crucial role in wound healing (1). Raising oxygen tension (pO_2) has shown to increase the rate of keratinocyte migration, collagen deposition and tensile strength (2,3). Systemic hyperbaric oxygen therapy (HBOT) and topical oxygen therapy (TOT) have been used to deliver oxygen to wounds (4–6). HBOT and TOT both have shown promising effects on wound revascularization and epithelialization. However, HBOT has higher costs associated and may have more side effects including ear discomfort and CNS toxicity (7). TOT is less expensive and can be performed at home (8). However, its ability to penetrate the skin is limited (9,10).

Recently developed topical aqueous oxygen emulsions (TOE) can overcome these limitations (9–11). TOE contains super-

saturated oxygen which can be delivered topically. The formulation is based on perfluorocarbon droplets being encapsulated within an aqueous continuous phase allowing slow release of oxygen over time. The oxygen solubility of the perfluorocarbon is relatively high, approximately twenty times greater than water; therefore, it has a high oxygen-carrying capacity (9,11). We have previously reported that TOE increased the healing rate in partial-thickness wounds in a porcine study *in vivo* (9). However, the mechanisms involved and molecular effects of the TOE on wound healing are still unknown.

Questions addressed

This study was to examine the molecular effects of the TOE in wound epithelialization, angiogenesis and granulation tissue formation.

國科會補助計畫衍生研發成果推廣資料表

日期:2013/10/17

國科會補助計畫	計畫名稱: 細胞核基質蛋白Matrin 3 (MATR3)在人類D型肝炎病毒RNA的合成過程中的重要性
	計畫主持人: 許國堂
	計畫編號: 101-2320-B-040-006- 學門領域: 寄生蟲學、醫事技術及實驗診斷
無研發成果推廣資料	

101 年度專題研究計畫研究成果彙整表

計畫主持人：許國堂		計畫編號：101-2320-B-040-006-					
計畫名稱：細胞核基質蛋白 Matrin 3 (MATR3) 在人類 D 型肝炎病毒 RNA 的合成過程中的重要性							
成果項目		量化			單位	備註 (質化說明：如數個計畫共同成果、成果列為該期刊之封面故事...等)	
		實際已達成數 (被接受或已發表)	預期總達成數 (含實際已達成數)	本計畫實際貢獻百分比			
國內	論文著作	期刊論文	0	0	50%	篇	
		研究報告/技術報告	0	0	100%		
		研討會論文	0	0	100%		
		專書	0	0	100%		
	專利	申請中件數	0	0	100%	件	
		已獲得件數	0	0	100%		
	技術移轉	件數	0	0	100%	件	
		權利金	0	0	100%	千元	
	參與計畫人力 (本國籍)	碩士生	0	0	100%	人次	
		博士生	0	0	100%		
		博士後研究員	0	0	100%		
		專任助理	0	0	100%		
國外	論文著作	期刊論文	1	2	50%	篇	
		研究報告/技術報告	0	0	100%		
		研討會論文	0	0	100%		
		專書	0	0	100%		章/本
	專利	申請中件數	0	0	100%	件	
		已獲得件數	0	0	100%		
	技術移轉	件數	0	0	100%	件	
		權利金	0	0	100%	千元	
	參與計畫人力 (外國籍)	碩士生	0	0	100%	人次	
		博士生	1	0	100%		
		博士後研究員	0	0	100%		
		專任助理	0	0	100%		

<p>其他成果 (無法以量化表達之成果如辦理學術活動、獲得獎項、重要國際合作、研究成果國際影響力及其他協助產業技術發展之具體效益事項等，請以文字敘述填列。)</p>	<p>無</p>
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	成果項目	量化	名稱或內容性質簡述
科 教 處 計 畫 加 填 項 目	測驗工具(含質性與量性)	0	
	課程/模組	0	
	電腦及網路系統或工具	0	
	教材	0	
	舉辦之活動/競賽	0	
	研討會/工作坊	0	
	電子報、網站	0	
	計畫成果推廣之參與(閱聽)人數	0	

國科會補助專題研究計畫成果報告自評表

請就研究內容與原計畫相符程度、達成預期目標情況、研究成果之學術或應用價值（簡要敘述成果所代表之意義、價值、影響或進一步發展之可能性）、是否適合在學術期刊發表或申請專利、主要發現或其他有關價值等，作一綜合評估。

1. 請就研究內容與原計畫相符程度、達成預期目標情況作一綜合評估

達成目標

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實驗失敗

因故實驗中斷

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說明：此計畫之後續實驗因為其複雜性甚高，需要資深的研究生來操作。當下資深的研究生已經畢業，而新的學生尚無能力執行後段複雜的研究。若能有三年的計畫來訓練和執行應該會有比較好的結果，只有壹年的計畫實在很不容易吸引研究生，尤其是在私立大學的招生相對為弱勢之下。

2. 研究成果在學術期刊發表或申請專利等情形：

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技轉： 已技轉 洽談中 無

其他：（以 100 字為限）

已發表 Hsiao YP, Yang JH, Wu WJ, Lin MH, Sheu* GT. (2013 Oct) E6 and E7 of human papillomavirus type 18 and UVB irradiation corporately regulate interleukin-6 and interleukin-8 expressions in basal cell carcinoma. *Exp Dermatol*. 22(10):672-674. doi: 10.1111/exd.12223, PMID: 24079741 (SCI) (IF=3.578 R/C=6/58=10.3%, DERMATOLOGY) (NSC-101-2320-B-040-006)

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如果我們可以辨識細胞內負責合成 HDV 病毒的 HDV 核酸聚合酶(RdRP)的工作羣，我們將有機會瞭解到 pol II 工作羣如何和 HDV RNA 進行作用，這對於醫學和生物學上都可以有相當的影響性。